

Distribution and Diversity of Seaweed Species in South Coastal Waters in Sri Lanka

Amal D Premarathna^{1*,2}, Kumara AMCP², Anura P Jayasooriya^{3,4}, Jayanetti DE^{2,5}, Ranjith B Adhikari⁶, Sarvananda L⁷, Amarakoon S⁸

¹Department of Veterinary Pathobiology, Faculty of Veterinary Medicine and Animal Science, University of Peradeniya, Peradeniya, Sri Lanka; ²Faculty of Fisheries & Marine Science, Ocean University of Sri Lanka, Tangalle, Sri Lanka; ³Department of Basic Veterinary Sciences, Faculty of Veterinary Medicine and Animal Science, University of Peradeniya, Peradeniya, Sri Lanka; ⁴Department of Basic Veterinary Sciences, Faculty of Medical Sciences, University of West Indies, Trinidad and Tobago, West Indies; ⁵NAQDA Freshwater Prawn Breeding Centre, Pambala, Sri Lanka; ⁶Department of Farm Animal Production and Health, Faculty of Veterinary Medicine and Animal Science, University of Peradeniya, Peradeniya, Sri Lanka; ⁷Department of Botany, Faculty of Sciences, University of Peradeniya, Peradeniya, Sri Lanka; ⁸Department of Animal Science, Faculty of Agricultural & Food Sciences, University of Manitoba, Winnipeg, Canada

ABSTRACT

Seaweeds are aquatic angiosperms, which are widespread in many coastal settings. Due to their various type of application, for instance, therapeutic agents, they are considered as one of the key environmental resources. In order to find out the diversity, abundance and pattern of species distribution of algae, a study was carried out in the three selected south coastal areas in the sand plain (Ahagama, Talpe and Koggala) in Sri Lanka. Quadrats method and three-line transects (horizontal & vertical) with 15 m length laid perpendicular from shore were adopted to collect data. Diversity was determined by Shannon-Wiener diversity index and quality of seawater (water temperature, pH, and salinity and dissolved oxygen) was also determined. Statistical analysis revealed significant difference in distribution and diversity between *Sargassum* and *Ulva* species in three study area. "Shannon-Wiener" diversity index value indicated a close relationship of the diversity in Koggala and Ahangama when compared to Talpe. It was also obvious that a positive correlation ($P < 0.05$) existed between *Sargassum ilicifolium* and *Ulva lactuca*. This study provides clear evidence that patterns and habitats of *Sargassum ilicifolium* and *Ulva lactuca* differ along the south coastline in Sri Lanka.

Keywords: Distribution; Quadrat; Seaweeds; *Sargassum sp*; *Ulva sp*

INTRODUCTION

Seaweeds are macroscopic, multicellular, benthic marine algae, and recognized as important marine natural resources. There are three major classes namely, Chlorophyceae, Rhodophyceae and Phaeophyceae [1]. However, their distribution is limited from the lower intertidal to the shallow subtidal zones of the marine environment. From them, an array of substances has been extracted and their applications are involved as stabilizers and stiffeners in food industry, precursors for cosmetics and biotechnology [2-4]. Recent studies conducted using seaweeds or their compounds have explored new avenues in medical advancement; bioactive compounds [5,6], as a CO₂ sink [7] and as bio-fuel [8]. Over the last few decades, marine natural products have been used as a leading compound for the discovery of drug in every part in the world [9].

In addition, they have been widely used as model organism to study biogeographic patterns and for testing of various ecological theories. Some of the seaweeds have been explored under rocks and in the intertidal habitat [10-12]. Presence of mudflats, estuaries, and coral reefs along the lagoons and rocky beaches provides an ideal habitat for sustainable growth of seaweeds in Sri Lanka [13].

The primary living cells on earth emerged from the sea and life has developed from the growth of mono-cellular algae [14]. Moreover, it's estimated about 90% of marine plant species are algae which contribute 40% global synthesis [15]. The Sri Lankan coastal water is rich in marine flora and variety of seaweeds has been identified [16]. The marine algae had been reported as early as the 19th century [17,18]. Further there are 440 taxa of marine algae belonging to 148 genera. Amongst the Sri Lankan seaweeds, *Sargassum* species

Correspondence to: Amal D Premarathna, Department of Veterinary Pathobiology, Faculty of Veterinary Medicine and Animal Science, University of Peradeniya, Peradeniya, Sri Lanka, Tel: +94775407879; E-mail: amaldharmapriya@gmail.com

Received: November 18, 2019; **Accepted:** January 06, 2020; **Published:** January 13, 2020

Citation: Premarathna AD, Kumara AMCP, Jayasooriya AP, Jayanetti DE, Adhikari RB, Sarvananda L, et al. (2020) Distribution and Diversity of Seaweed Species in South Coastal Waters in Sri Lanka. J Oceanogr Mar Res 7:196. doi: 10.35248/2572-3103.19.7.196

Copyright: © 2019 Premarathna AD, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

are found in most of the coastal areas [19]. Furthermore, species such as *Laminaria*, *Fucus*, *Ascophyllum* and *Tubinaria* are also found in the Sri Lankan waters. Since 1960 to 1970 alginic acid has been extracted from seaweeds [20], and the cultivation of seaweeds too has been introduced [21]. A further study on the various aspects of macro algae is needed to diversify and find the richness of the species.

The main focus of this study is on the diversity and distribution of seaweeds along the south-coast of Sri Lanka. However, information regarding the identities and diversity of the economically important seaweeds species are lacking in Sri Lanka. Such information can provide a baseline for future complex ecological studies in this area and further to manage and explore seaweeds.

METHODS

Description of the study area

The study site selected for the sampling is the south coastal area with the geographical platform of Ahangama (N 05° 58.006' E 080° 22.482'), Koggala (N 05° 59.390' E 080° 33.121') and Talpe (N 05° 59.792' E 080° 16.898') in Sri Lanka and research was conducted in 2015. Accordingly, nine stations were selected to perform sampling of seaweeds from this geographical setting. The sites varied in distance from one another ranging from 7.2 km to 4.0 km (Figure 1).

Collection of data and samples

The zonation profile of the study area and distribution pattern of the macrophytes was observed physically by placing three transects perpendicular to the shore [22]. All the seaweed samples were taken by snorkeling from shallow waters (0-2 m in depth) along the subtidal area. At each site, different types of macrophyte specimens were collected manually by hand and using a knife during the low tide. All the samples were stored in pre-labeled plastic bags while macro algae were collected in plastic pots containing 5% neutral

formalin. These specimens were identified using the identification guide [20] and voucher specimens and herbarium sheets were prepared. Furthermore, the seaweeds were authenticated at the "National Herbarium of the Peradeniya Botanical Garden in Sri Lanka" and specimens were deposited for future references.

Sampling site and quadrats method

To estimate the cover of each seaweed species in each location, we used representative sites and random quadrats of 90 × 90 cm² and subdivided into 81 square units of 10 × 10 cm² with thick nylon line. The presence or absence of the species were recorded for each unit. Transect lines marked in 1m increments were placed at random at different sites. They were used as guides for placing the subdivided quadrats contiguously along the transect. These transects were aligned perpendicular to the coastline. Each extended outwards from the rocky shore to the sand bottom at a distance varying from 9-15m, depending on the inclination of the shore. Snorkeling divers were used to assess the vertical distribution and abundance of the seaweed species. 8100 cm² of quadrat (90 × 90 cm²) was used to estimate the percentage of cover by the dominant benthic seaweeds.

When sampling, the time and depth at each quadrat were recorded and the depth was corrected according to the tide level. Three depth ranges were sampled at all sites. Abundance of each algae group was expressed as number of individuals of that group/m² as a percentage of the total number of algae individuals of all species / m² (Figure 2).

Statistical analysis

Graph Pad Prism Version 4.03 for Windows (Graph Pad Software, San Diego, CA, USA) and SPSS were used for the statistical analysis. All data were obtained in order to show the seaweed distribution and their relative abundance (%). The percentage of taxa was estimated for the communities at each depth site. To test the differences in species and depths, 09 quadrats were randomly

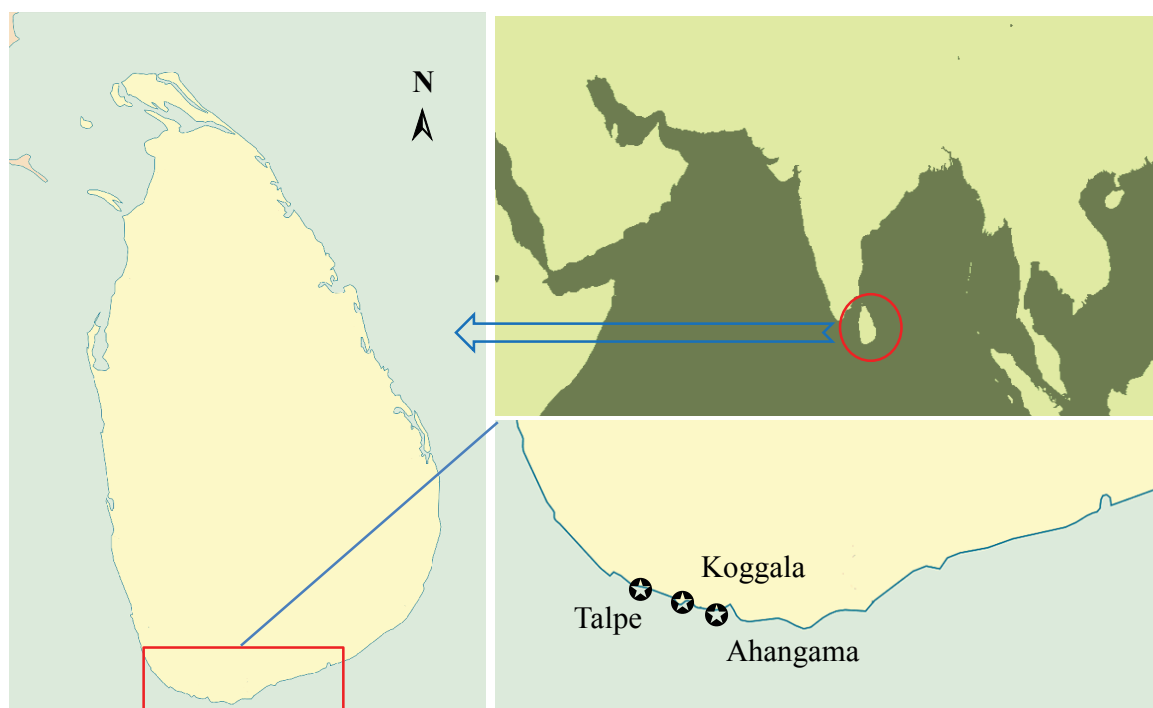


Figure 1: Map of the south coastal region in Sri Lanka, showing the sampling sites from where seaweeds were collected.

sub-sampled from the data sets and divided in three depth intervals. At each site, overall mean percentage covers of each taxon were calculated using one-way and two-way analysis of variance. Multiple comparisons between the significant levels of interactions of the variables were done by Tukey's method and values were expressed as the Mean \pm SE and $P < 0.05$ considered significant.

RESULTS

Shannon-Wiener diversity index values indicated that the diversity of Koggala was closely related to the diversity of Ahangama whereas the diversity of Talpe was not closely related to the diversity of Ahangama and Koggala coastal areas (Figure 3).

A statistically significant difference was found between seaweed distributions in Talpe, Koggala and Ahangama coastal areas (Table 1).

A statistically significant relationship was found between abundance of *Sargassum* species and *Ulva* species. *Sargassum*

ilicifolium dominates the lower intertidal zone when compared with other seaweeds species. *Ulva lactuca* occurs closer to the shore. There was significant difference with temperature, salinity and pH, dissolved oxygen and wave within the chosen study site (Figure 4).

Out of eight species, *Ulva lactuca* was the most common and widely spread in Talpe coastal area, where as *Sargassum ilicifolium* is widely spread in Ahangama coast. A statistical relationship ($P < 0.05$) was found to exist between relative abundance of *Sargassum ilicifolium* and *Ulva lactuca*. *Sargassum ilicifolium* appears in Koggala coastal area water with high dissolved oxygen content. *Ulva lactuca* species appears in water with low dissolved oxygen content and a quiet environment when compared with *Sargassum* species distribution (Figure 5).

Two algae species, *Sargassum ilicifolium* and *Ulva lactuca* were selected to characterize the intertidal zone and explore the macrophyte patterns in the shallow subtidal regions. The brown

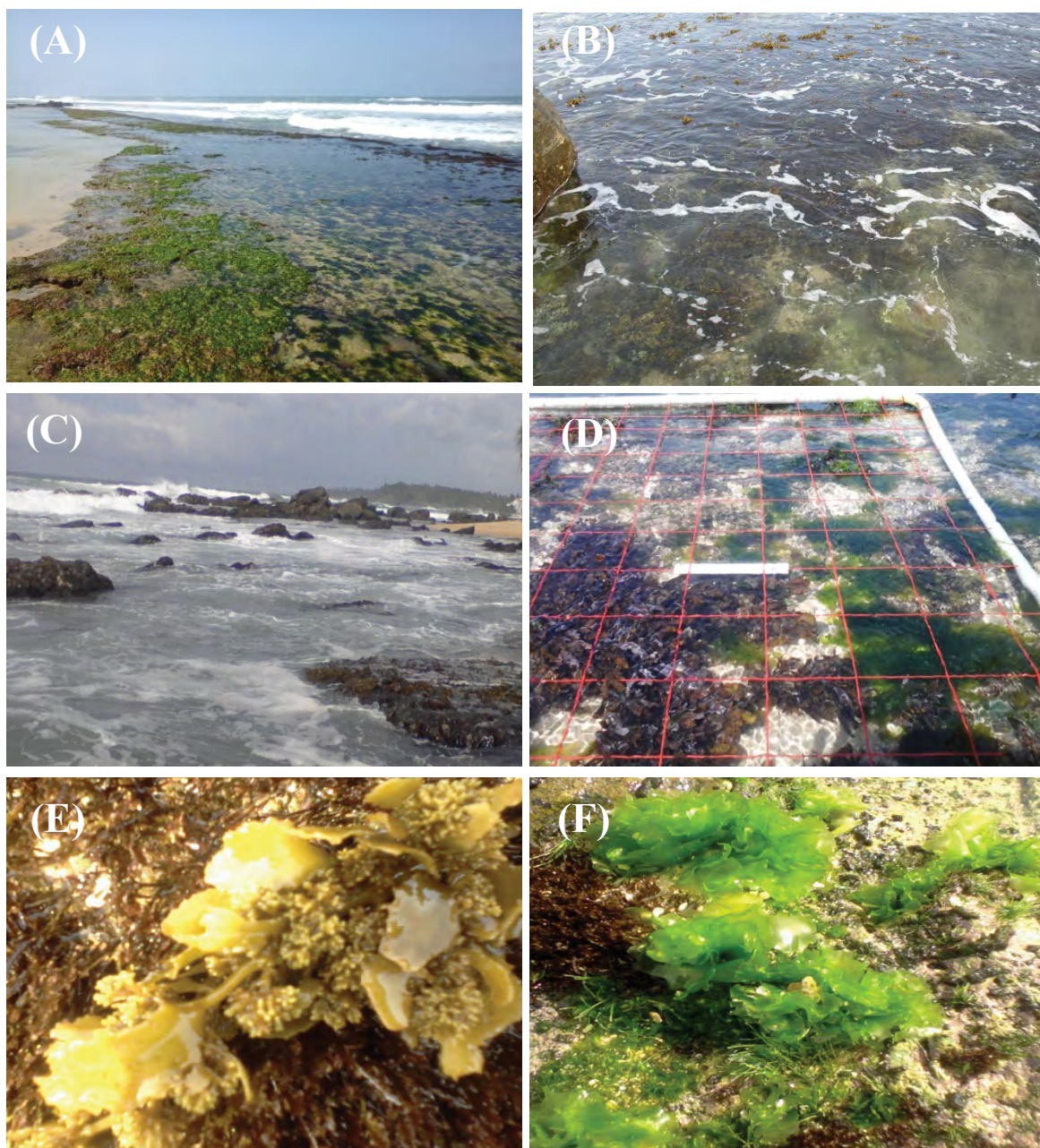


Figure 2: Seaweeds of coastal waters of the southwest coast of Sri Lanka. A) Talpe, B) Koggala, C) Ahangama, D) *Sargassum* sp. and *Ulva* sp. monitor to characterize the intertidal zone and estimate the cover of each algal species in using quadrats of $90 \times 90 \text{ cm}^2$, E) *Sargassum ilicifolium*, F) *Ulva lactuca*.

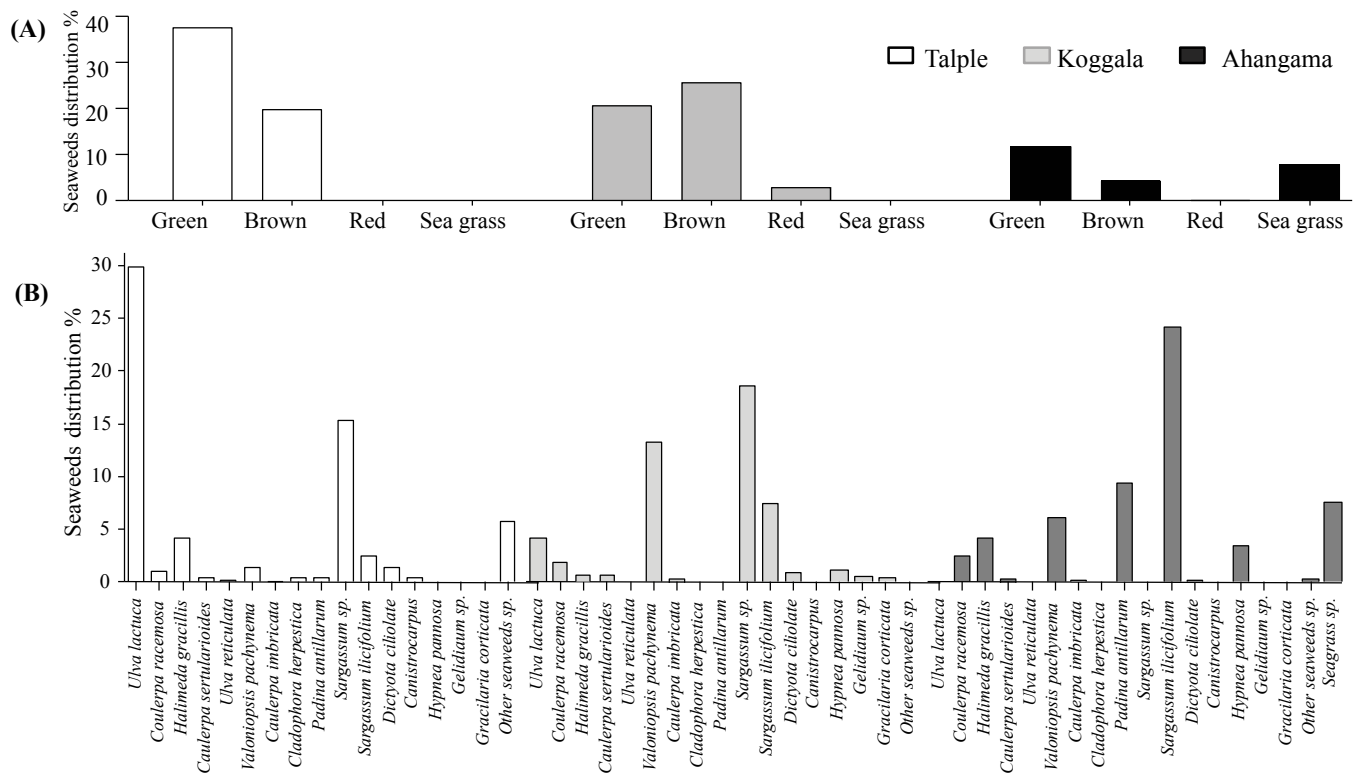


Figure 3: Bar chart showing the distribution of seaweeds A) Distribution of Green, Brown, and Red seaweeds. B) The Distribution of seaweed species. Percentages are (specific species cover)/(total seaweed cover) × 100.

Table 1: Species list of seaweeds at Talpe, Koggala and Ahangama coastal in Sri Lanka.

Algae Species Name	Talpe	Koggala	Ahangama
<i>Ulva Lactuca</i>	+	+	-
<i>Caulerpa racemosa</i>	+	+	+
<i>Halimeda gracillis</i>	+	+	+
<i>Caulerpa sertularioides</i>	+	+	+
<i>Ulva reticulata</i>	+	-	-
<i>Valoniopsis pachynema</i>	+	+	+
<i>Caulerpa imbricata</i>	+	+	+
<i>Cladophora herpestica</i>	+	-	-
<i>Padina antillarum</i>	+	-	+
<i>Sargassum sp.</i>	+	+	-
<i>Sargassum ilicifolium</i>	+	+	+
<i>Dictyota ciliolate</i>	+	+	+
<i>Canistrocarpus sp.</i>	+	-	-
<i>Hypnea pannosa</i>	-	+	+
<i>Gelidium sp.</i>	-	+	-
<i>Gracilaria corticata</i>	-	+	-

+ Present; - Absent

seaweed of *Sargassum ilicifolium* and green seaweed of *Ulva lactuca* be the important groups in abundance. They ranged respectively from 13-66% and 27-68% of total algae. Considering from the ecological and economic view, *Ulva lactuca* could be an important green seaweed in the Talpe area. Besides, the importance of these aquatic inter-tidal macrophytes for fishery resources and overall ecosystem processes should not be over looked in this coastal area.

DISCUSSION

Horizontal distribution of the benthos

We recognized the existence of benthic communities occupying the same habitat but they were in different depth zones. The algal community seem to be dominated by turf algae (mostly green seaweeds) at shallowest depths and the brown seaweed *Sargassum ilicifolium* were found at deeper levels. Different patterns were found at different sites, demonstrating that different spatially discrete factors may be acting on each rocky shore. Brown seaweeds *Sargassum ilicifolium* have more cover ($19.59 \pm 9.129\%$) than other seaweeds at Ahangama. In this area *Ulva lactuca* species are not present. This *Sargassum ilicifolium* species has been shown to dominate the brown seaweeds of many regions in Sri Lanka. At the Talpe sites a green seaweed *Ulva lactuca* ($17.67 \pm 5.930\%$) community dominated and *Sargassum ilicifolium* community coverage ($2.497 \pm 1.679\%$) was small in this area. Koggala sites were normally covered by *Sargassum ilicifolium* ($7.106 \pm 2.078\%$) and *Ulva lactuca* ($3.992 \pm 2.644\%$).

Vertical distribution of the benthos

Our analysis indicates that the distribution of taxa showed a clear vertical differentiation. Vertical distribution of species on rocky shores varies due to many factors such as hydrodynamics [23], turbidity [24], temperature [25], wave exposure [26-28], slope [29], light intensity [30], grazing activities [31-33], and predation [34]. Ahangama area has been colonized by *Sargassum ilicifolium* which is distributed in lower intertidal zone. The site in Ahangama located at a narrow depth range of (0.5 -1.5 m) housed sea grasses and brown algae (*Sargassum ilicifolium*, *Padina antillarum* and *Dictyota*

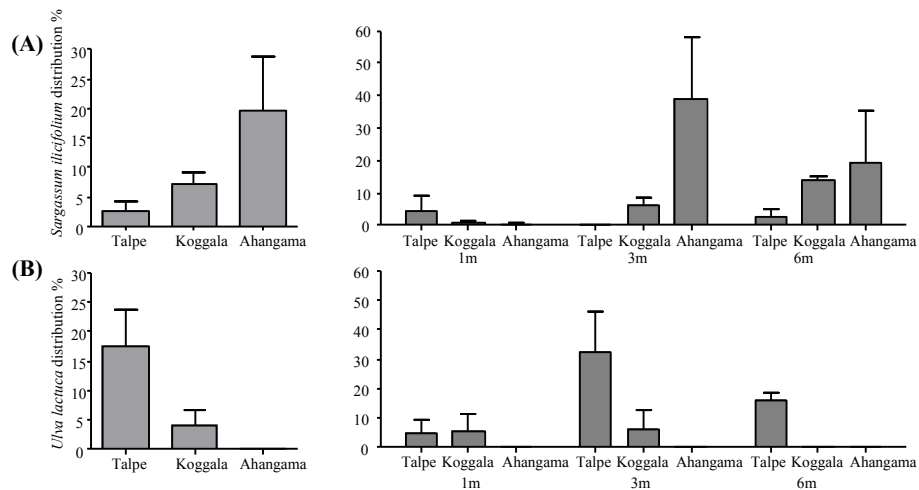


Figure 4: *Sargassum sp.* and *Ulva sp.* distribution in south coastal area. A) Distribution of *Sargassum ilicifolium* B) Distribution of *Ulva lactuca*. The left-side figures in 4A and 4B show the total amount of *Sargassum ilicifolium* and *Ulva lactuca* respectively. Percentages are (specific species cover)/(total seaweed cover) $\times 100$.

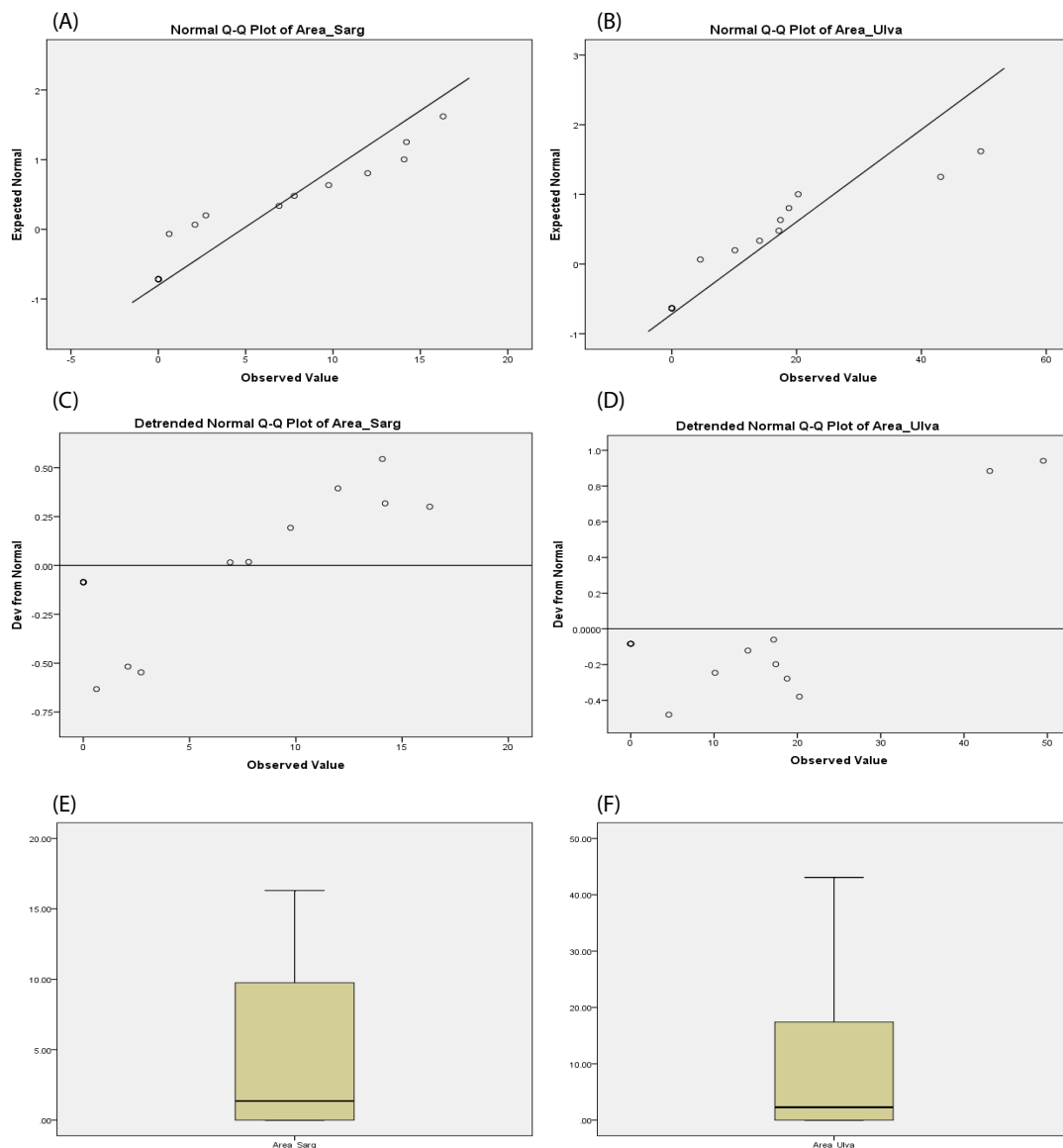


Figure 5: Normal Q-Q Plot and Detrended Normal Q-Q Plot of Average percentage of seaweed distribution. A) Normal Q-Q Plot of Average percentage of *Sargassum ilicifolium*, B) Normal Q-Q Plot of Average percentage of *Ulva lactuca*, C) Detrended Normal Q-Q Plot of *Sargassum ilicifolium* distribution, D) Detrended Normal Q-Q Plot of *Ulva lactuca* distribution, E) Box Plot of *Sargassum ilicifolium* distribution, F) Box Plot of *Ulva lactuca* distribution.

ciliolate), red algae (*Hypnea pannosa*), green algae (*Halimeda gracilis*, *Valoniopsis pachynema*, *Caulerpa sertularioides*, *Coulerpa racemosa*, *Caulerpa imbricate*). The intertidal zone is the area of the marine shoreline that affects the amount of incident light and so the development of an algal community [35]. The algae bed slope was gentle and shallow; plenty of light was available, so growth rates of macro-algae were high [36]. At Koggala the depth and high turbidity reduce light intensity and so reduce growth of algal species.

The brown algae (*Sargassum sp.*, *Padina antillarum*, *Sargassum ilicifolium*, *Canistrocarpus* and *Dictyota ciliolate*), green algae (*Halimeda gracilis*, *Coulerpa racemosa*, *Valoniopsis pachynema*, *Caulerpa sertularioides*, *Ulva lactuca*, *Ulva reticulate*, *Cladophora herpestica*, *Caulerpa imbricate*) are favoured as substrates and are among the dominant algae species along the Talpe coastal area. The rough sea obstructed sampling at Koggala and extension of rocky shore only up to 10 m limit the collection of samples at Talpe.

Human activities are intense in coastal areas and marine communities are subjected to pressure from a multitude of stressors, resulting in large scale changes in the abundance and distribution of species [37,38]. Ahangama algae bed is situated very close by the fisheries harbor and anglers can utilize this area for their fishing activities every day. The brown algae (*Sargassum sp.*, *Sargassum ilicifolium* and *Dictyota ciliolate*) red algae (*Gracilaria corticata*, *Gelidium sp.*, *Hypnea pannosa*) and green algae (*Halimeda gracilis*, *Valoniopsis pachynema*, *Caulerpa sertularioides*, *Ulva lactuca*, *Caulerpa imbricate*) are favoured as substrates and are among the dominant algae along the Koggala coastal area. The brown algae *Sargassum ilicifolium* and green algae *Ulva lactuca* were found as substrates since they are among the dominant algae forms along the Ahnagama and Talpe coastal area respectively.

CONCLUSION

This study provides evidence that the most abundantly distributed seaweed species in this area are brown and green seaweeds. However, brown seaweed namely; *Sargassum* species are found to be the commonest in the area most exposed to sea waves with high dissolved oxygen contents, whereas green seaweed namely; *Ulva* species dominates in the shallow depths. Further we observed changing patterns of distribution in communities of seaweeds along the depth gradient; in fact, the highest diversity is obvious close to the coast. Density of *Sargassum ilicifolium* species was higher in Ahangama compared with Talpe and Koggala. It was also obvious that a positive correlation ($P < 0.05$) existed between *Sargassum ilicifolium* and *Ulva lactuca*. It is pronounced that there is a marked difference of respective seaweed communities in Ahangama and Talpe. However, further investigations are essential to establish assertive correlations among seaweed community, structure and distribution in these locations.

AVAILABILITY OF DATA AND MATERIALS

The data and materials are contained within the paper. Samples and data collections were obtained from the south coastal algae bed in Sri Lanka.

COMPETING INTERESTS

The author(s) declare that they have no competing interests.

FUNDING

This research did not receive any specific grant from funding agencies in the public, commercial, or non-profit sectors.

AUTHOR'S CONTRIBUTIONS

AD conceived the study, concept, and design and conducted sample collection; analyzed, interpreted results. AD, LS, AMCP and RB analyzed data and manuscripts preparation. SA and AP interpretation data and contributed with critical revision of the manuscript. AD and DE contributed with sample collection and analysis. All authors read and approved the final manuscript.

ACKNOWLEDGEMENTS

The authors wish to thank Faculty of Fisheries & Marine Science, Ocean University of Sri Lanka for providing us with a pleasant environment to complete this research project. Support given by Mr. KA Wijesekera by editing the manuscripts is also highly appreciated.

REFERENCES

1. Trono Jr GC, Ganzon-Fortes ET. Philippines seaweeds, Technology and livelihood center PCPM Certificate of registration No. 1988.
2. Aneiros A, Garateix A. Bioactive peptides from marine sources: pharmacological properties and isolation procedures. *J Chromatogr B*. 2004;803(1):41-53.
3. Jeeva S, Kiruba S. Bio-remediating and bio mediating potential of seaweeds. National Seminar on Marine Resources: Sustainable Utilization and Conservation, Organized by Department of Plant Biology and Biotechnology, St. Mary's College, Thoothukudi, 38p, 2009.
4. Wiencke C, Amsler CD. Seaweeds and their communities in polar regions. In *Seaweed Biology*, Springer, Berlin, Heidelberg. 2012;265-291.
5. Baby VA, Appavoo MR, Huxley VAJ. Antibacterial effects of *Ulva* faciatia extracts on nosocomial pathogen *Pseudomonas aeruginosa*. *J Basic Appl Biol*. 2012;5:204-208.
6. Ranaheva TH, Premarathna AD, Jayasooriya LJPAP, Wijesundara RRMKK, Rajapakse RPVJ. In-vitro anti-cancer and cytotoxic properties of aqueous seaweed extracts on BHK and HeLa cell lines. In *Proceedings of the International Research Symposium on Pure and Applied Sciences (IRSPAS 2016)*, Faculty of Science, University of Kelaniya, Sri Lanka. 2016:pp:07.
7. Muraoka D. Seaweed resources as a source of carbon fixation. *Bulletin-Fisheries Research Agency Japan*, 2004;pp:59-64.
8. Bastianoni S, Coppola F, Tiezzi E, Colacevich A, Borghini F, Focardia S. Biofuel potential production from the Orbetello lagoon macroalgae: A comparison with sunflower feedstock. *Biomass and Bioenergy*. 2008;32(7):619-628.
9. Premarathna AD, Ranaheva TH, Wijesekera SK, Wijesundara RRMKK, Jayasooriya AP, Wijewardana V, et al. Wound healing properties of aqueous extracts of *Sargassum ilicifolium*: An in vitro assay. *Wound Med*. 2019;24(1):1-7.
10. Murray SN, Littler MM. Analysis of seaweed communities in a disturbed rocky intertidal environment near Whites Point, Los Angeles, Calif., USA. *Eleventh International Seaweed Symposium*. Springer, Dordrecht, 1984.
11. Bolton JJ, Leliaert F, De Clerck O, Anderson RJ, Stegenga H, Engledow HE, et al. Where is the western limit of the tropical Indian

- Ocean seaweed flora? An analysis of intertidal seaweed biogeography on the east coast of South Africa. *Mar Biol.* 2004;144(1):51-59.
12. Prathep A. Spatial and temporal variations in diversity and percentage cover of macroalgae at Sirinart Marine National Park, Phuket Province, Thailand. *Science Asia.* 2005;31:225-233.
 13. Premarathna AD, Sarvananda L, Jayasoorya AP, Amarakoon S. A Review on Pathogenic Diseases on Corals Associated Risk Factors and Possible Devastations in Future in the Globe. *J Mar Sci Res Dev.* 2019;9:269.
 14. Dhargalkar VK, Pereira N. Seaweed: promising plant of the millennium. *Sci Cult.* 2005;71(3-4):60-66.
 15. Andersen RA. Diversity of eukaryotic algae. *Biodivers Conserv.* 1992;1(4):267-292.
 16. Borgesen F. Some marine algae from Ceylon. *Ceylon J Sci Sect A Bot.* 1936;12:57-96.
 17. BARTON ES. List of marine algae collected by Professor Herdman, at Ceylon, in 1902, with a note on the fructification of Halimeda. WA Herdman, Report to the Government of Ceylon on the pearl oyster fisheries in the Gulf of Manaar. Part I. London: The Royal Society. 1903; p:163-167.
 18. Baldwin DA. Infants' contribution to the achievement of joint reference. *Child Dev.* 1991;62(5):875-890.
 19. Durairatnam M. Contribution to the study of the marine algae of Ceylon. *Bulletin of the Fisheries Research Station Ceylon.* 1961;10:1-181.
 20. Arumugam I, Sivapalan A, Theivendirajah K. Preliminary studies on the alginic acid and agar contents of some marine algae. *J Nat Sci Coun Sri Lanka.* 1985;13(2):197-212.
 21. Coppejans E, Leliaert F, Dargent O, Gunasekara R, De Clerck O. Sri Lankan seaweeds: Methodologies and field guide to the dominant species. *Belgian Development Cooperation.* 2009;6:265.
 22. English SS, Wilkinson CC, Baker VV. Survey manual for tropical marine resources. *Aust Inst Mar Sci.* 1997;Pp:390.
 23. Barkai A, Branch GM. Contrasts between the benthic communities of subtidal hard substrata at Marcus and Malgas islands: A case of alternative stable states?. *South African J Mar Sci.* 1988;7(1):117-137.
 24. Saiz-Salinas JJ, Urkiaga-Alberdi J. Faunal responses to turbidity in a man-modified bay (Bilbao, Spain). *Mar Env Res.* 1999;47(4):331-347.
 25. Menge BA, Sutherland JP. Community regulation: variation in disturbance, competition, and predation in relation to environmental stress and recruitment. *The American Naturalist.* 1987;130(5):730-757.
 26. Huston MA. Patterns of species diversity on coral reefs. *Annu Rev Ecol Syst.* 1985;16(1):149-177.
 27. Castro CB, Echeverria CA, Pires DO, Mascarenhas BJ, Freitas SG. Distribuicao De Cnidaria E Echinodermata No Infralitoral De Costoes Rochosos De Arraial Do Cabo, Rio De Janeiro, Brasil. *Anais da Academia Brasileira de Ciencias.* 1993;65(4):458-458.
 28. Ferreira CE, Gonçaves JE, Coutinho R. Community structure of fishes and habitat complexity on a tropical rocky shore. *Env Biol Fishes.* 2001;61(4):353-369.
 29. REVIE W. Coral populations on reef slopes and their major controls. *Mar Ecol Prog Ser.* 1982;7:83-115.
 30. Figueiredo MDO. Colonization and growth of crustose coralline algae in Abrolhos, Brazil. In *Proceedings of the 8th international coral reef symposium, Panama.* 1997;1:689-694.
 31. Jones NS, Kain JM. Subtidal algal colonization following the removal of Echinus. *Helgolander Wissenschaftliche Meeresuntersuchungen.* 1967;15(1-4):460-466.
 32. Menge BA, Lubchenco J. Community organization in temperate and tropical rocky intertidal habitats: prey refuges in relation to consumer pressure gradients. *Ecological Monographs.* 1981;51(4):429-450.
 33. Patricio F, Dearborn JH. Community structure of macroinvertebrates inhabiting the rocky subtidal zone in the Gulf of Maine: seasonal and bathymetric distribution. *Mar Ecol Prog Ser.* 1989;57:147-161.
 34. Hardwick-Witman MN. Biological consequences of ice rafting in a New England salt marsh community. *J Exper Mar Biol Ecol.* 1985;87(3):283-298.
 35. Earll RC. *Sublittoral ecology: The ecology of the shallow sublittoral benthos.* Oxford University Press, UK. 1983.
 36. Cole KM, Sheath RG. (Eds.). *Biology of the red algae.* Cambridge University Press, USA. 1990.
 37. Ranahewa TH, Gunasekara AJM, Premarathna AD, Karunarathna SC, Jayamanne SC. A Comparative Study on the Diversity of Seagrass Species in Selected Areas of Puttalam Lagoon in Sri Lanka. *J Oceanogr Mar Res.* 2018;6(3):2.
 38. Worm B, Chapman AR. Interference competition among two intertidal seaweeds: *Chondrus crispus* strongly affects survival of *Fucus evanescens* recruits. *Mar Ecol Prog Ser.* 1996;145:297-301.

This article was originally published in a special issue, **Marine Biology** handled by Editor(s). Dr. Jeffrey Craig Bailey, Associate Professor, Department of Biology & Marine Biology The University of North Carolina, USA.